

NANOPORE SEQUENCING BASECALLING

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Data Structure

Data



Events



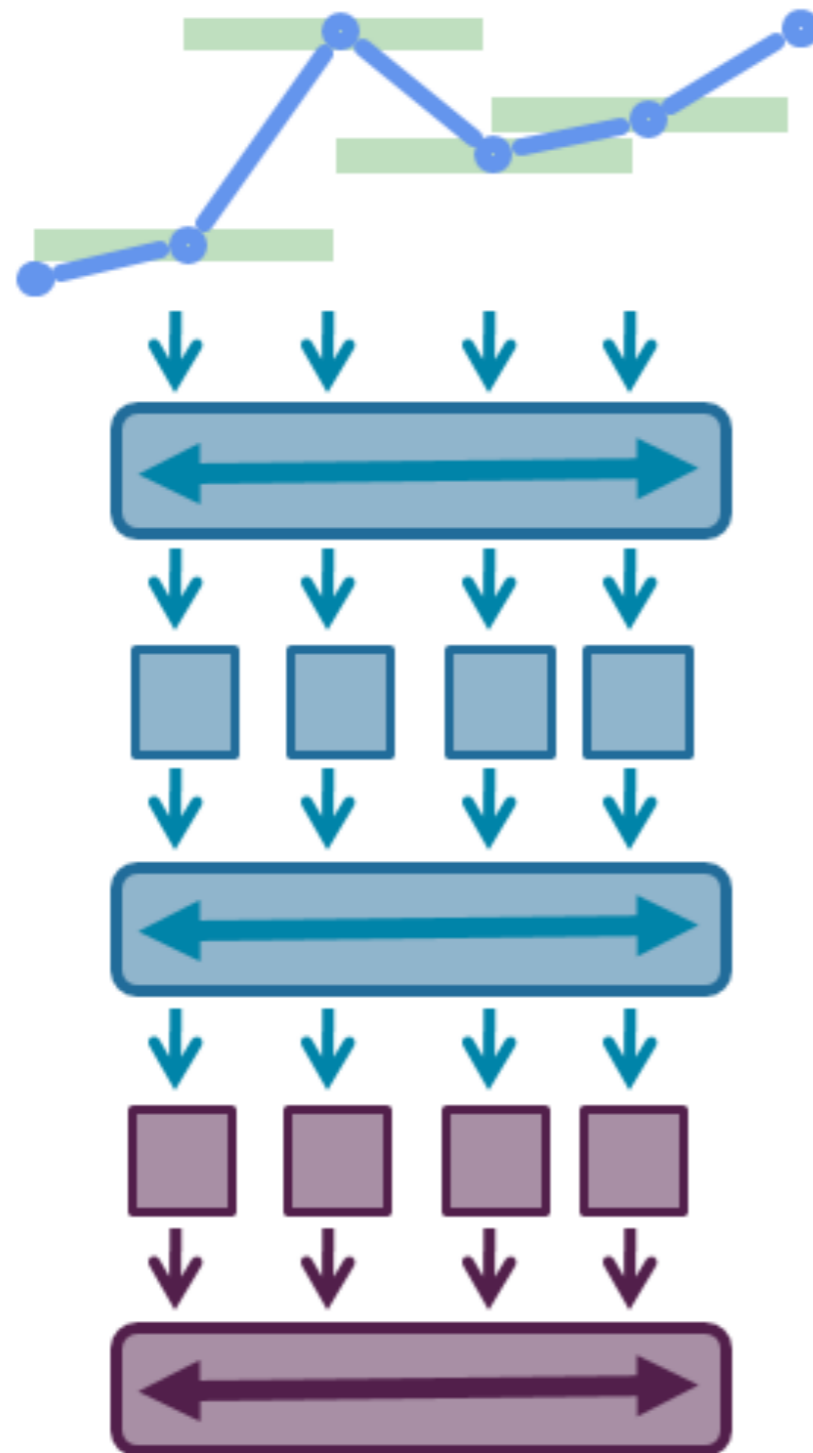
Sequence

```
ONT1 CCGACTCCGGTTACCCGCGTTGATTTGCTGGGGCAGGGCCG
      |||:|||||
REF   CCGACTCCGGTTACCAGCGTTGATTTGCTGGGGCAGGGCCG
```

Basecalled

- * Raw data - a direct measurement of the changes in ionic current as a DNA/RNA strand passes through the pore. These measurements are recorded by the MinKNOW software. MinKNOW also processes the signal into reads, each read corresponding to a single strand of DNA/RNA.
- * Basecalling - the raw signal is processed into segments with information about current level, noise level and duration, which correspond to the movement of the DNA through the pore. These data are further processed by the basecalling algorithm to generate the base sequence of the read.

Basecalling - RNN



Distributions learned from squiggle training data

Bidirectional information flow (BLSTM layer)

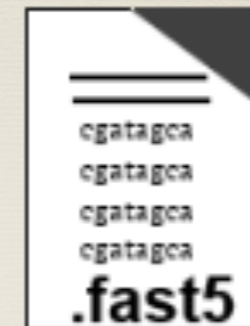
Processing layer

Bidirectional information flow (BLSTM layer)

Multi-base prediction

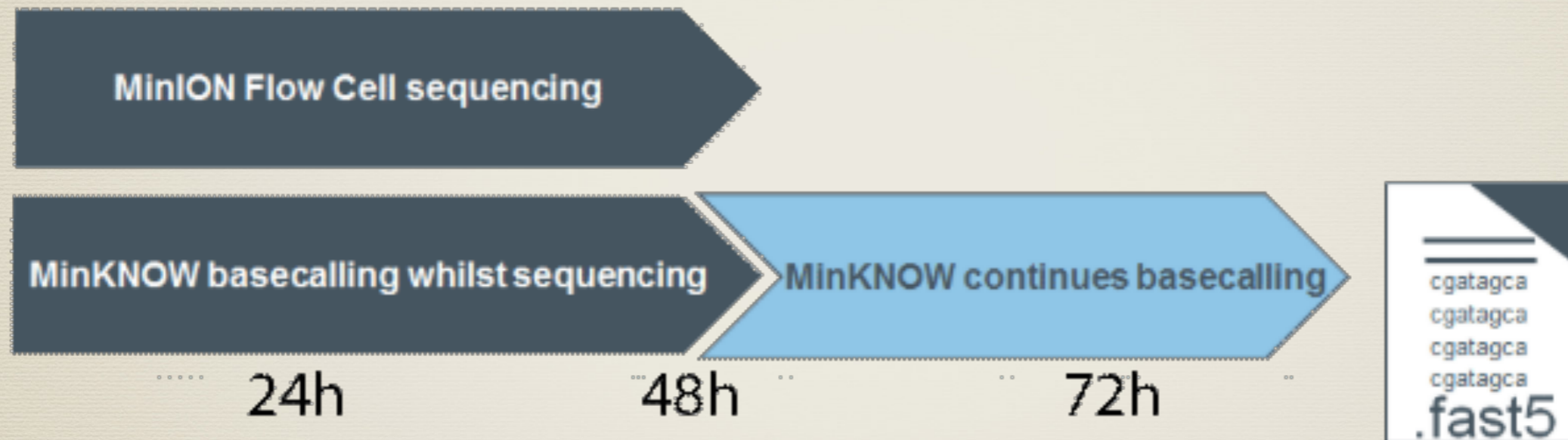
Decode to sequence

Live basecalling with MinKNOW



MinKNOW basecalling modes

Catch-up mode



Keep-up mode - requires higher computational power usually not available with a laptop

Oxford Nanopore basecallers

| Basecaller | Algorithm |
|-----------------------|--|
| MinKNOW basecaller | Production basecaller; uses a neural network. The algorithm is identical to the one used by Albacore, but may be a version behind |
| Albacore | Production basecaller; uses a neural network. Currently available as an executable, or as source code for members of the Developer group |
| Nanonet | Research basecaller; uses the latest research algorithm. Not actively supported |
| Scrappie | Research basecaller; uses a neural network with a 'transducer' model, which allows the basecaller to resolve long homopolymers. Not actively supported |

Basecalling components

- * Event detection
- * Segmentation
- * iD basecalling



Read file location

- * Windows
 - * :\\data\\reads on the SSD
- * Mac OS X
 - * /Library/MinKNOW/data
- * Linux
 - * /var/lib/MinKNOW/data



File location and name

The reads are stored in `data_folder/reads`, where "data_folder" is the location that the user sets up during MinKNOW installation. This location does not change during autoupdates, but if the software is reinstalled, the installer will ask for the location.

The filename for a read follows a hardcoded pattern:

```
<output_reads_dir>/tmp/<batch_number>/  
<data_set>_ch<channel_number>_read<read_number>.fast5.tmp
```

and if the partially completed read is not otherwise rejected, when it is completed:

```
<data_folder>/reads/<basecall_status>/<batch_number>/  
<data_set>_ch<channel_number>_read<read_number>_<classification>.fast5
```


File content

```
{attributes: file_version}  
-UniqueGlobalKey/  
  |-tracking_id/{attributes: asic_id, asic_id_eeprom, asic_temp,  
    device_id, exp_script_hash, exp_script_name, exp_script_purpose, exp_start_time,  
    flow_cell_id, heatsink_temp, hostname, protocol_run_id, protocols_version_name,  
    run_id, version, version_name}  
  |-channel_id/{attributes: channel_number, digitisation, offset, range,  
    sampling_rate}  
  |-context_tags/{attributes: set when the experiment is configured}  
-Raw/  
  |-Reads/  
    |-Read_42/{attributes: start_time, duration, read_number, start_mux,  
      read_id, median_before}  
    |-Signal{samples}  
  |
```


Albacore

- * Albacore is a software that provides an entry point to the Oxford Nanopore basecalling algorithms.
- * It can be run from the command line on Windows and multiple unix-like platforms.
- * A selection of configuration files allow basecalling DNA libraries made with our current range of sequencing kits and Flow Cells.



Albacore

System requirements

- 4 GB RAM plus 1 GB per worker thread for 1D basecalling
- 4 GB RAM plus 2 GB per worker thread for 1D2 and 2D basecalling
- Administrator access for installation
- ~100 Mb of drive space for installation, minimum 512 GB storage space for basecalled read files (1 TB recommended)
- When starting with a .fast5 file that only has raw data in it, the file size will increase approx. 5 times



Albacore

- * read_fast5_basecaller.py
- * main script written in python
- * an entry gate to ONT basecalling



read_fast5_basecaller.py -l

| Flow cell | Seq kit | | Flow cell | Seq kit |
|------------------------|----------------------|--|------------------------|----------------------|
| FLO-MIN _{I06} | SQK-LSK ₂ | | FLO-MIN _{I07} | SQK-RNA ₀ |
| FLO-MIN _{I06} | SQK-NSK ₀ | | FLO-MIN _{I07} | SQK-RAD ₀ |
| FLO-MIN _{I06} | SQK-RAD ₀ | | FLO-MIN _{I07} | SQK-RAD ₀ |
| FLO-MIN _{I06} | SQK-RLI ₀ | | FLO-MIN _{I07} | SQK-RLI ₀ |
| FLO-MIN _{I06} | SQK-LWP ₀ | | FLO-MIN _{I07} | SQK-LWP ₀ |
| FLO-MIN _{I06} | SQK-RAS ₂ | | FLO-MIN _{I07} | SQK-RAS ₂ |
| FLO-MIN _{I06} | SQK-LSK ₁ | | FLO-MIN _{I07} | SQK-LSK ₁ |
| FLO-MIN _{I06} | VSK-VBK ₀ | | FLO-MIN _{I07} | VSK-VBK ₀ |
| FLO-MIN _{I06} | SQK-RBK ₀ | | FLO-MIN _{I07} | SQK-DCS ₁ |
| FLO-MIN _{I06} | SQK-RLB ₀ | | FLO-MIN _{I07} | SQK-PCS ₁ |
| FLO-MIN _{I06} | SQK-LWB ₀ | | FLO-MIN _{I07} | SQK-RBK ₀ |
| FLO-MIN _{I06} | SQK-RAB ₂ | | FLO-MIN _{I07} | SQK-RLB ₀ |
| FLO-MIN _{I06} | SQK-RNA ₀ | | FLO-MIN _{I07} | SQK-LWB ₀ |
| | | | FLO-MIN _{I07} | SQK-RAB ₂ |
| | | | FLO-MIN _{I07} | SQK-LSK ₃ |

Basecalling example

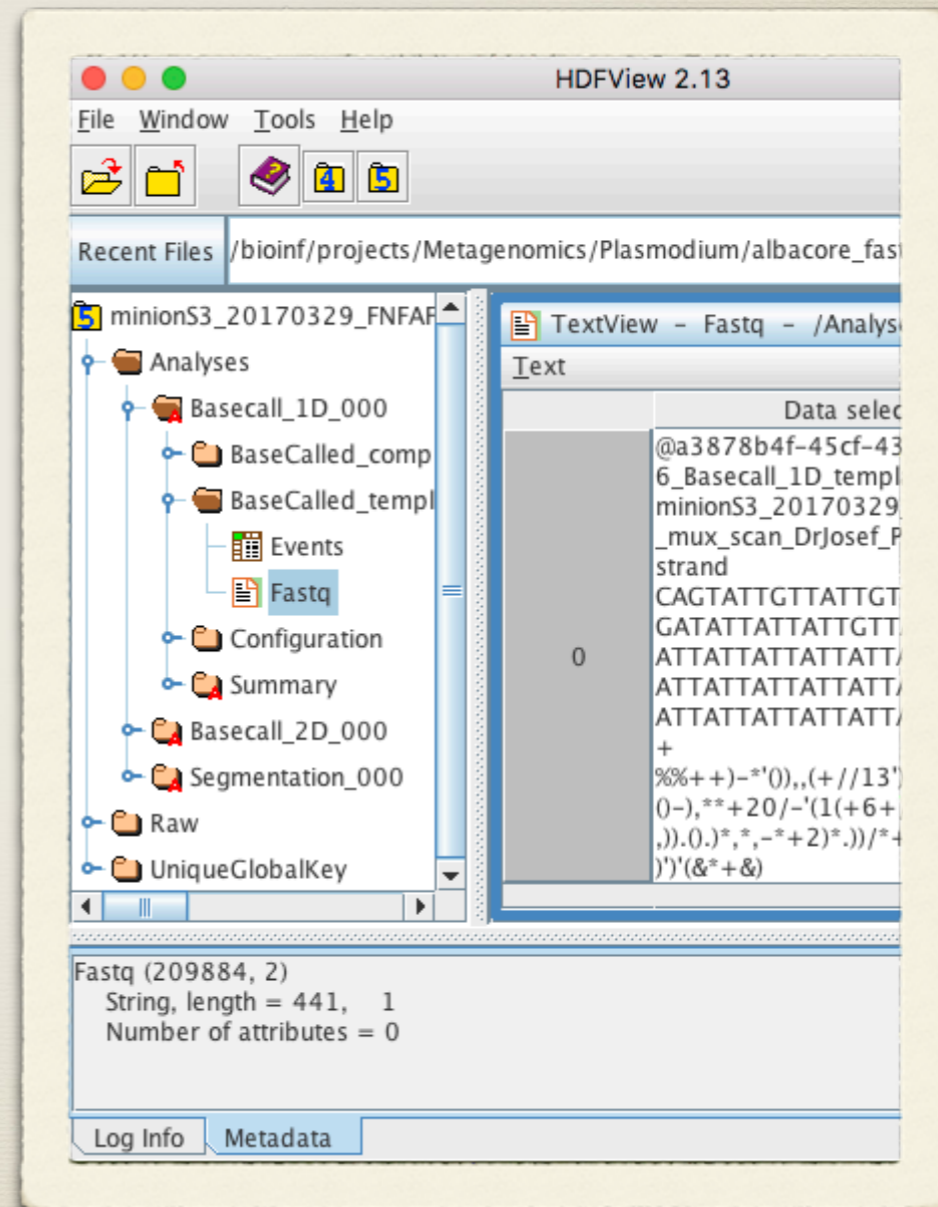
```
read_fast5_basecaller.py -i 20170519_I639_I70519_run -t 2  
-s 20170519_I639_I70519_run -o fastq,fast5 -r  
-f FLO-MIN106 -k SQK-LSK208
```



MinION data formats

FAST5

- * The raw data is stored as binary files in HDF5 standard
- * HDFView allows quick look at the raw data files
- * <https://support.hdfgroup.org/products/java/release/download.html#bin>



MinION data formats

FASTQ

```
@SRR001666.1 071112_SLXA-EAS1_s_7:5:1:817:345 length=36  
GGGTGATGGCCGCTGCCGATGGCGTCAAATCCCACC  
+  
IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII9IG9IC
```

Line 1 begins with a '@' character and is followed by a sequence identifier and an optional description

Line 2 is the raw sequence letters.

Line 3 begins with a '+' character and is optionally followed by the same sequence identifier (and any description) again.

Line 4 encodes the quality values for the sequence in Line 2, and must contain the same number of symbols as letters in the sequence.

MinION data formats

FASTQ

$$Q = -10 \log_{10} p$$

p = probability that the corresponding base call is incorrect

| ASCII | p | Q |
|-------|--------|----|
| ! | 1 | 0 |
|) | 0.1 | 10 |
| 3 | 0.01 | 20 |
| = | 0.001 | 30 |
| H | 0.0001 | 40 |
| ~ | | 93 |

!"#\$%&'()* +,-./0123 456789:;<= >?@ABCDEFGHI I

MinION data formats

FASTA

Very simple format but it may contain quite a bit in formation on the sequence.

Used by many software including BLAST and NanoPipe

```
>SRR001666.1 071112_SLXA-EAS1_s_7:5:1:817:345 length=36  
GGTGATGGCCGCTGCCGATGGCGTCAAATCCCACC
```

